

Client: Stephanie Green,
Ethos Environmental Planning



ADAS
Spring Lodge
172 Chester Road
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WAS 0AR

Tel: 01159 229249
Email: Helen.Rees@adas.co.uk
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Sample ID: ADAS-3587 Condition on Receipt: Good Volume: Passed
Client Identifier: Wilson's Pearce's Farm Description: pond water samples in preservative
Date of Receipt: 05/05/2022 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control ¹	2 of 2	Real Time PCR	09/05/2022
Degradation Control ⁵	Within Limits	Real Time PCR	09/05/2022
Great Crested Newt*	0 of 12 (GCN negative)	Real Time PCR	09/05/2022
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) ⁶	4 of 4	Real Time PCR	As above for GCN

Report Prepared by: Dr Helen Rees Report Issued by: Dr Ben Maddison

Signed:



Signed:



Position: Director: Biotechnology Position: MD: Biotechnology

Date of preparation: 10/05/2022 Date of issue: 10/05/2022

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

* If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.

¹ Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

⁵ No degradation is expected within time frame of kit preparation, sample collection and analysis.

⁶ Additional positive controls (10⁻¹, 10⁻², 10⁻³ ng/μL) are also routinely run, results not shown here.

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Sample ID: ADAS-3608 Condition on Receipt: Medium Sediment Volume: Passed

Client Identifier: Pearce's Farm Description: pond water samples in preservative

Date of Receipt: 05/05/2022 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	2 of 2	Real Time PCR	09/05/2022
Degradation Control [‡]	Within Limits	Real Time PCR	09/05/2022
Great Crested Newt*	0 of 12 (GCN negative)	Real Time PCR	09/05/2022
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) [§]	4 of 4	Real Time PCR	As above for GCN
Report Prepared by:	Dr Helen Rees	Report Issued by:	Dr Ben Maddison

Signed:



Signed:



Position: Director: Biotechnology

Position: MD: Biotechnology

Date of preparation: 10/05/2022

Date of issue: 10/05/2022

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

* If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.

[†] Recorded as the number of positive replicate reactions at expected C_i value. If the expected C_i value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

[‡] No degradation is expected within time frame of kit preparation, sample collection and analysis.

[§] Additional positive controls (10⁻², 10⁻³, 10⁻⁴ ng/μL) are also routinely run, results not shown here.

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Sample ID: ADAS-3618 Condition on Receipt: Good Volume: Passed
Client Identifier: Prior's Farm Goldfish Description: pond water samples in preservative
Date of Receipt: 05/05/2022 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	2 of 2	Real Time PCR	09/05/2022
Degradation Control [§]	Within Limits	Real Time PCR	09/05/2022
Great Crested Newt*	0 of 12 (GCN negative)	Real Time PCR	09/05/2022
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) [‡]	4 of 4	Real Time PCR	As above for GCN

Report Prepared by: Dr Helen Rees Report Issued by: Dr Ben Maddison

Signed:



Signed:



Position: Director: Biotechnology Position: MD: Biotechnology

Date of preparation: 10/05/2022 Date of issue: 10/05/2022

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

** If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.*

[†] Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

[§] No degradation is expected within time frame of kit preparation, sample collection and analysis.

[‡] Additional positive controls (10⁻¹, 10⁻², 10⁻³ ng/μL) are also routinely run, results not shown here.

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Sample ID: ADAS-3593 Condition on Receipt: Good Volume: Passed

Client Identifier: Priors Farm, Corp Pond Description: pond water samples in preservative

Date of Receipt: 05/05/2022 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	2 of 2	Real Time PCR	09/05/2022
Degradation Control [§]	Evidence of degradation	Real Time PCR	09/05/2022
Great Crested Newt*	Indeterminate	Real Time PCR	09/05/2022
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) [‡]	4 of 4	Real Time PCR	As above for GCN
Report Prepared by:	Dr Helen Rees	Report Issued by:	Dr Ben Maddison
Signed:		Signed:	
Position:	Director: Biotechnology	Position:	MD: Biotechnology
Date of preparation:	10/05/2022	Date of issue:	10/05/2022

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

* If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.

[†] Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

[§] No degradation is expected within time frame of kit preparation, sample collection and analysis.

[‡] Additional positive controls (10⁻¹, 10⁻², 10⁻³ ng/μL) are also routinely run, results not shown here.

Client:

Priors Farm, Stephanie Green, Ethos Environmental Planning
1040079-SG Ethos, Priors Farm, version 1



RSK ADAS Ltd
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Sample ID: ADAS-9664

Client Identifier: P3

Grid references/coordinates: Not Supplied

Description: pond water samples in preservative

Condition on Receipt: Low Sediment

Date of Receipt : 28/05/2025

Volume: Passed

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	2 of 2	Real Time PCR	06/09/2025
Degradation Control [‡]	Within limits	Real Time PCR	06/09/2025
Great Crested Newt*	1 of 12 (positive)	Real Time PCR	06/09/2025
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) [§]	4 of 4	Real Time PCR	As above for GCN
Report Prepared by:	Dr Helen Rees	Report Issued by:	Dr Ben Maddison
Signed:		Signed:	
Position:	Director: Biotechnology	Position:	MD: Biotechnology
Date of preparation:	09/06/2025	Date of issue:	09/06/2025

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

* If all PCR controls and extraction blanks give the expected results a sample is considered: negative for GCN if all of the replicates are negative; positive for GCN if one or more of the replicates are positive.

[†] Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not

[‡] No degradation is expected within time frame of kit preparation, sample collection and analysis.

[§] Additional positive controls (10⁻¹, 10⁻², 10⁻³ ng/μL) are also routinely run, results not shown here.

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1040079-SG-Ethos, Priors Farm, version 1



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Sample ID: ADAS-9579

Client Identifier: P26

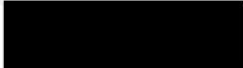
Grid references/coordinates: Not Supplied

Description: pond water samples in preservative

Condition on Receipt: Good

Date of Receipt : 10/06/2025

Volume: Passed

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	2 of 2	Real Time PCR	13/06/2025
Degradation Control [§]	Within limits	Real Time PCR	13/06/2025
Great Crested Newt*	0 of 12 (negative)	Real Time PCR	13/06/2025
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) [#]	4 of 4	Real Time PCR	As above for GCN
Report Prepared by:	Dr Helen Rees	Report Issued by:	Dr Ben Maddison
Signed:		Signed:	
Position:	Director: Biotechnology	Position:	MD: Biotechnology
Date of preparation:	13/06/2025	Date of issue:	13/06/2025

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

* If all PCR controls and extraction blanks give the expected results a sample is considered: negative for GCN if all of the replicates are negative; positive for GCN if one or more of the replicates are positive.

[†] Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not

[§] No degradation is expected within time frame of kit preparation, sample collection and analysis.

[#] Additional positive controls (10⁻¹, 10⁻², 10⁻³ ng/μL) are also routinely run, results not shown here.